

Chemical Composition and Antibacterial Activity of *Artemisia herba-alba* and *Mentha pulegium* Essential Oils

Houda Sbayou¹, Bouchra Ababou², Khadija Boukachabine², Angeles Manresa³, Khalid Zerouali⁴ and Souad Amghar¹

1. Laboratory of Agrofood and Health, Faculty of Sciences and Technologies, University Hassan I^{er}, Settat Km 3, B.P: 577, Morocco

2. Laboratory of Environmental Sciences and Development, Faculty of Sciences and Technologies, University Hassan I^{er}, Settat Km 3, B.P: 577, Morocco

3. Unity of Microbiology, Faculty of Pharmacy, University of Barcelona, Av. Joan XXIII s/n, 08028 Barcelona, Spain

4. Laboratory of Microbiology, CHU Ibn Rochd. 1, Street of Hospitals, Morocco

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Abstract: The chemical composition of essential oils obtained from *Artemisia herba-alba* and *Mentha pulegium* were determined. The essential oils were analyzed by gas chromatography coupled with mass spectrometry (GC/MS). Their antibacterial activity was studied *in vitro* against three standard strains: *E. coli* ATCC 25922, *Staphylococcus aureus* ATCC 29213, *Pseudomonas aeruginosa* ATCC 27853, and five clinical strains: *Enterobacter cloacae*, *Staphylococcus aureus*, *Pseudomonas pyocyaneque*, *Enterococcus faecium*, and *E. coli*. Nineteen constituents were identified in *A. herba-alba* essential oil representing 99.57% of the total composition. The major component was α -thujone (59.07%). The bacterial strains were inhibited at concentrations ranging from 1.25 μ L/mL to 5 μ L/mL and killed at concentrations ranging from 1.25 μ L/mL to 10 μ L/mL. *M. pulegium* resulted in the identification of eighteen constituents representing 99.48% of the total composition. The main component was pulegone (78.07%). The minimal inhibitory (MIC) and bactericidal (MBC) concentrations were ranging from 1.25 μ L/mL to 2.5 μ L/mL.

Key words: *Artemisia herba-alba*, *Mentha pulegium*, GC/MS (gas chromatography-mass spectrometry), antibacterial activity.

1. Introduction

Therapy of bacterial infection is mainly based on the use of antibiotics. The widely and sometimes the inappropriate use of these agents, led to the development of multidrug resistant strains, hence the importance of directing research towards new ways especially toward herbal medicine that have always been a source of inspiration for new drugs.

Essential oils are volatile, natural, complex compounds characterized by a strong odor and are formed by aromatic plants as secondary metabolites [1]. They are widely used in medicine as constituents

of different medical products, in the food industry as flavouring additives and also in cosmetics as fragrances [2].

Indeed, several studies have confirmed the antigenotoxic [3, 4], antibacterial [5, 6] and antifungal effects [7] of some essential oils and their components.

Artemisia herba-alba is a medicinal and aromatic dwarf shrub, that commonly grown in Mediterranean basin [8]. In Morocco, this plant is found in the Oriental regions, the Eastern Rif, the Middle Atlas, the High Atlas and the Saharan Anti-Atlas [9]. It is used extensively in traditional medicine to treat helminthiasis, diabetes mellitus and other

Corresponding author: Souad Amghar, Ph.D., professor, research field: microbiology. E-mail: eamghar@gmail.com.

conditions such as jaundice [10]. Also, the antihyperglycaemic [11], antimicrobial [12], antioxidant, antispasmodic, anti-venom, nematocidal, anthelmintic, anti-leishmanial, neurological, pesticidal and inhibitor activities of this plant have previously been reported [13].

Mentha pulegium is one of the *Mentha* species known as pennyroyal, a native herb of Asia and near East [14]. In Morocco, this plant grows in wet areas [9]. It has been traditionally used as antiseptic for treatment of cold, sinusitis, cholera, food poisoning, bronchitis and tuberculosis [15].

The aim of this study was to determine the chemical composition of essential oils of *A. herba-alba* and *M. pulegium* grown in Morocco and to investigate antibacterial activities against some clinical strains that exhibit multidrug resistance to commonly used antibiotics.

2. Material and Methods

2.1 Essential Oils

Essential oils used in this study were provided by Santis Company. They are extracted by steam distillation from flowers, leaves and stems (Table 1).

2.2 Microorganisms

The tested strains included the following bacteria: three standard strains: *Escherichia coli* ATCC 25922, *Staphylococcus aureus* ATCC 29213, *Pseudomonas aeruginosa* ATCC 27853 (Microbiology Laboratory, Faculty of Pharmacy, University of Barcelona, Spain) and five clinical isolates *Enterobacter cloacae*, *Staphylococcus aureus*, *Pseudomonas pyocyanique*, *Enterococcus faecium* and *Escherichia coli* (Microbiology Laboratory, CHU Ibn Rochd, Casablanca, Morocco).

2.3 Gas Chromatography-Mass Spectrometry Analysis

The chromatographic analysis of essential oils was performed with a gas chromatograph (Trace GC Ultra) coupled to a mass spectrometer (Polaris Q ion

trap MS), with a VB-5 capillary column (methylpolysiloxane with 5% phenyl; 30 m × 0.25 mm; film thickness 0.25 µm). Fragmentation was performed by electron impact at 70 eV. Helium (1.4 mL/min) was used as carrier gas. Split-type injector was heated to a temperature of 200 °C. The volume injected was 1 µL. The column was initially maintained at a temperature of 40 °C for 2 min, increased to 180 °C at a rate of 4 °C/min, and finally raised to 300 °C for 2 min at 20 °C/min.

2.4 Disc Diffusion Method

The tested as described previously [16]. Sterile filter paper disc (6 mm diameter) were impregnated with 10 µL of essential oil and transferred into the Luria Bertoni Agar present in Petri dishes, which had previously seeded by spreading 1 mL of bacterial suspension adjusted to 10⁶ CFU/mL. Standard antibiotics amoxicillin (25 µg/disk) were used as positive control. After incubation at 37 °C during 24 h, the diameters of inhibition zones were measured in millimeters. Tests were carried out in triplicate.

2.5 Determination of MIC and MBC

MIC was determined in this work by the method of macro-broth dilution [17]. A serial of dilution of essential oil ranging from 20 µL/mL to 0.15 µL/mL were prepared in test tubes containing Broth Luria Bertoni medium with 0.15% Agar [18]. Each tube was inoculated with the same volume of bacterial suspension adjusted to 10⁶ CFU/mL. The tubes were then incubated at 37 °C for 18 h. MIC values were defined as the lowest concentration of the EO at which the absence of growth was recorded. Controls of medium with either microorganisms or the essential oil alone were included. From tubes where it was no trouble, aliquots of 10 µL were inoculated on Muller Hinton Agar medium. The MBC was the lowest concentration that gives no subculture. Each assay was repeated thrice.

3. Results and Discussion

3.1 Chemical Composition of Essential Oils

Analysis of the chemical composition of *A.*

herba-alba essential oil by gas chromatography coupled with mass spectrometry (GC/MS) revealed the presence of nineteen compounds representing 99.57% of the total composition (Table 2).

Table 1 Region and period of collection of each plant studied.

Plant species	Region of collection	Period of collection
<i>Artemisia herba-alba</i>	Taroudant: Southeast Morocco	April-June 2009
<i>Mentha pulegium</i>	Taounate: Northeast Morocco	April-July 2009

Table 2 Chemical composition (%) of two essential oils from *Aremisia herba-alba* and *Mentha pulegium*.

Compounds	Percentage of compounds (%)	
	<i>Artemisia herba-alba</i>	<i>Mentha pulegium</i>
α -Pinene	0.68	0.41
Artemisia triene	3.75	
Sabinene	3.05	
2-Cyclohexen-1-ol, 1-methyl-4-(1-methylethyl)-, cis-	0.93	
ζ -Terpinene	0.27	
α -Thujone	59.07	
2,4-Hexadiene, 2,3-dimethyl-	11.73	
α -CAMPHOLENE ALDEHYDE	12.71	
Bicyclo[3.1.1]hept-2-ene-2-ethanol, 6,6-dimethyl- (CAS)	0.72	
2- α -PINENE	0.25	
Patchoulane	0.30	
trans-2-Caren-4-ol	0.13	
α -Murolene	0.16	
GERMACRENE-D	0.44	0.19
1H-Cycloprop[e]azulene, decahydro-1,1,7-trimethyl-4-methylene-, [1aR-(1a α ,4a α ,7 α ,7a α ,7b α)]-	0.14	
1,1,3,3,5,5,7,7,9,9,11,11-DODECAMETHYL-HEXASILOXANE	5.24	13.37
3-Carene		0.29
(+)-Camphene		0.80
Cyclohexene,4-ethyl-3-ethylidene-4,8-		0.50
Bis(2-propylamino)-2,6-dichloro-1,5- naphoquinone		0.14
3-Cyclopentene-1-ethanol, 2,2,4-trimethyl-		0.35
1-MENTHONE		0.74
Isomenthone		0.36
Cyclohexanone, 5-methyl-2-(1-methylethenyl)-, trans		1.45
Pulegone		78.07
α -Cedrol		0.49
Bicyclo[5.2.0]nonane, 2-methylene-4,8,8-trimethyl-4-vinyl-		0.80
α -Caryophyllene		1.19
10aH-2,12a-Methano-1H,4H-cyclopropa[5,6][1,3]dioxolo[2',3']cyclopenta[1',2':9,10]cyclodeca[1,2-d][1,3]dioxin-15-ol, 1a,2,7a,13,14,14a-hexahydro-1,1,6,6,9,9,11,13-octamethyl-,acetate,[1aR-a α ,2 α ,7a α ,7bR*,10a α ,12a α ,13 α ,14a α ,15S*)]-		0.13
10aH-2,12a-Methano-1H,4H-cyclopropa[5,6][1,3]dioxolo[2',3']cyclopenta[1',2':9,10]cyclodeca[1,2-d][1,3]dioxin-15-ol, 1a,2,7a,13,14,14a-hexahydro-1,1,6,6,9,9,11,13-octamethyl-, [1aR-(1a α ,2 α ,7a α ,7bS*,10a α ,12a α ,13 α ,14a α ,15R*)]-		0.20
Total	99.57%	99.48%

Among these compounds, five of them can be considered as the main constituents: α -thujone (59.07%); campholene aldehyde (12.71%); 2,4-Hexadiene, 2,3-dimethyl- (11.73%); Artemisia triene (3.75%) and Sabinene (3.05%). The α -thujone was also identified as the majority constituents of the essential oil of Matmata in Tunisia (44%) [19] and Jordan (16%) [20].

The chemical composition of *Artemisia herba-alba* essential oil of Taroudant is vastly different from that of M'sila (Algeria), which is dominated by camphor (19.4%) trans-pinocarveol (16.9%) chrysanthenone (15.8) and β -thujone (15%) [21]. Previous studies have shown that camphor was the main component of the *Artemisia herba-alba* of Algeria, Spain and Israel with a percentage between 15% and 68% [22-24].

The constituents of *Mentha pulegium* essential oil from Taounat are listed in Table 2. Chromatographic analyzes have identified eighteen compounds representing 99.48% of the total composition. The essential oil of *M. pulegium* is characterized by the presence of the pulegone as the main component with a percentage of 78.07%.

These results are similar to most of the work already done in Morocco [22-24]. Also, the work undertaken by Snoussi, et al. [25] and Hajlaoui, et al. [26] in Tunisia showed that pulegone was the major compound of *M. pulegium* with concentrations 44.27% and 61.11%, respectively. While work of Mahboubi, et al. [27] in Iran as well as those of Derwich, et al. [28] in Morocco highlighted another

chemotype whose major compounds are piperitone and piperitenone with low levels of pulegone. In addition to pulegone (43.3% to 87.3%), Beghidji, et al. [29] found in different sources of Algeria, a chemotype of *M. pulegium* characterized by its richness in monoterpene (α and β -pinene, camphene, sabinene, α -terpinene and myrcene).

The chemical composition of *Artemisia herba-alba* and *Mentha pulegium* essential oils shows a large interspecies variability, due to climatic and soil variations, to the vegetative cycle, and to seasonal variation.

3.2 Antibacterial Activity

The *in vitro* antimicrobial activity of *A. herba-alba* and *M. pulegium* essential oils against microorganisms was qualitatively and quantitatively assessed by the diameter of inhibition zone, MIC and MBC values.

To determine the MIC and MBC, the authors adopted the method of broth dilution using 0.15% agar to ensure the homogeneity of the oil-water mixture [33]. The diameter of inhibition zone, MIC and MBC results are shown in Tables 3 and 4.

The data indicated that the oils exhibited varying levels of antimicrobial activity against the investigated bacteria. With the exception of *Pseudomonas* strains that shows resistance to the bactericidal and bacteriostatic action of *A. herba-alba* and *M. pulegium* essential oils, all other bacteria are inhibited at concentrations ranging from 1.25 μ L/mL to 5 μ L/mL for essential oil of *A. herba-alba*, and from 1.25

Table 3 Diameter of inhibition zone in (mm).

	Microorganism species	Inhibition zone diameter in (mm)		Positive control (AMX)
		<i>A. herba-alba</i>	<i>M. pulegium</i>	
Gram–	<i>E. coli</i> ATCC 25922	16 $\pm$ 0	15 $\pm$ 0	25 $\pm$ 0
	<i>E. coli clinique</i>	17 $\pm$ 0	12.5 $\pm$ 0.7	0 $\pm$ 0
	<i>Ps. aeruginosa</i> ATCC 27853	0 $\pm$ 0	0 $\pm$ 0	0 $\pm$ 0
	<i>Ps. pyocyaneque</i>	8 $\pm$ 0	0 $\pm$ 0	0 $\pm$ 0
	<i>Enterobacter cloacae</i>	13 $\pm$ 0	11 $\pm$ 0	12 $\pm$ 0
Gram+	<i>St. aureus clinique</i>	11 $\pm$ 0	12 $\pm$ 0	12 $\pm$ 0
	<i>St. aureus</i> ATCC 29213	13 $\pm$ 0	15 $\pm$ 0	16 $\pm$ 0
	<i>Enterococcus faecium</i>	12 $\pm$ 0	12 $\pm$ 0	18 $\pm$ 0

Table 4 MIC and MBC of essential oils.

Microorganism species	MIC ($\mu\text{L/mL}$)		MBC ($\mu\text{L/mL}$)		MBC/MIC	
	<i>A. herba-alba</i>	<i>M. pulegium</i>	<i>A. herba-alba</i>	<i>M. pulegium</i>	<i>A. herba-alba</i>	<i>M. pulegium</i>
<i>E. coli</i> ATCC 25922	1.25	1.25	1.25	1.25	1	1
<i>E. coli clinique</i>	2.5	1.25	2.5	1.25	1	1
Gram– <i>Ps. aeruginosa</i> ATCC 27853	>20	>20	>20	>20		
<i>Ps. pyocyaneque</i>	>20	>20	>20	>20		
<i>Enterobacter cloacae</i>	1.25	2.5	1.25	2.5	1	1
<i>St. aureus clinique</i>	2.5	1.25	5	1.25	2	1
Gram+ <i>St. aureus</i> ATCC 29213	2.5	1.25	5	1.25	2	1
<i>Enterococcus faecium</i>	5	2.5	10	2.5	2	1

$\mu\text{L/mL}$ to 2.5 $\mu\text{L/mL}$ for the essential oil of *M. pulegium*. And killed at concentrations ranging from 1.25 $\mu\text{L/mL}$ to 10 $\mu\text{L/mL}$ for essential oil of *A. herba-alba* and from 1.25 $\mu\text{L/mL}$ to 2.5 $\mu\text{L/mL}$ for the essential oil of *M. pulegium*.

Ps. aeruginosa which proved resistant to the antibacterial effect of the essential oils tested is known by a high level of intrinsic resistance to virtually all known antimicrobial compounds including essential oils [34, 35]. This resistance seems to be related with the nature of the outer membrane which is composed of lipopolysaccharides that form an impermeable barrier to hydrophobic compounds [36, 37], but this was not true about *A. herba-alba* essential oil which showed a strong antibacterial effect on Gram– bacteria.

Among the gram-positive bacteria, *Enterococcus faecium* was less sensitive to the action of the essential oils tested with MIC = 5 $\mu\text{L/mL}$ for *A. herba-alba* essential oil and MIC = 2.5 $\mu\text{L/mL}$ for *M. pulegium* essential oil, while *E. coli* ATCC 25922 was more sensitive with MIC = 1.25 $\mu\text{L/mL}$ for both essential oils tested.

The MBC/MIC ratio is used to classify antibiotics according to their characters as bactericides (close to 1) or bacteriostatic (greater than 4) [38]. Our results showed that *A. herba-alba* and *M. pulegium* essential oils has a bactericidal activity against all bacteria tested excepted *Pseudomonas*. The presence of pulegone in *M. pulegium* essential oil and α -thujone in *A. herba-alba* essential oil may be responsible for their antibacterial activity. Indeed, it has reported that

pulegone play an important role in antibacterial activity [39, 40]. However, Other *Artemisia* oils rich in camphor and 1,8-cineole were previously demonstrated to have potent antimicrobial activities in vitro [41, 42]. Thus, it is difficult to attribute the activity of a complex mixture to a single or particular constituent. Major or trace compounds might give rise to the antimicrobial activity exhibited. Possible synergistic and antagonistic effect of compounds in the oil should also be taken into consideration.

4. Conclusions

In this work, we studied the chemical composition and antibacterial activity of the essential oil of *Artemisia herba-alba* from Taroudant and *Mentha pulegium* from Taounat. Chemical analysis by GC/MS identified respectively nineteen and eighteen constituents. The α -thujone (59.07%) was the major component of *A. herba-alba* while *M. pulegium* was dominated by pulegone (78.07%). The results obtained in this study show that the essential oils tested in vitro have significant activity against most bacteria tested.

References

- [1] F. Bakkali, S. Averbeck, D. Averbeck, M. Idaomar, Biological effects of essential oils—A review, Food and Chemical Toxicology 46 (2008) 446-475.
- [2] M.M. Cowan, Plant products as antimicrobial agent, Clin. Microbiol. Rev. 12 (1999) 564-582.
- [3] N. Mezzoug, A. Elhadri, A. Dallouh, S. Amkiss, N.S. Skali, J. Abrini, et al., Investigation of the mutagenic and antimutagenic effects of *origanum compactum* essential

- oil and some of its constituents, *Mutat. Res* 629 (2007) 100-110.
- [4] F. Bakkali, S. Averbeck, D. Averbeck, M. Idaomar, Biological effects of essential oils: A review, *Food Chem. Toxicol* 46 (2008) 446-475.
 - [5] H. Oumzil, S. Ghouami, M. Rhajaoui, A. Ildrissi, S. Fkih-tetouani, M. Faïd, et al., Antibacterial and antifungal activity of essential oils of *Mentha suaveolens*, *Phytother. Res* 16 (2002) 727-731.
 - [6] C.F. Bagamboula, M. Uyttendacle, J. Debevere, Inhibitory effect of thyme and basil essential oils, carvacrol, thymol, estragol, linalool and p-cymene towards *Shigella sonnei* and *S. flexneri*, *Food Microbiol* 21 (2004) 33-42.
 - [7] M.E. Ajjour, B. Satrani, M. Ghanmi, A. Aafi, A. Farah, M. Rahouti, et al., Antifungal activity of essential oils of *Thymus bleicherians* Pomel and *Thymus capitatus* (L.) Hoffm & Link against wood decay fungi, *Biotechnol. Agron. Soc. Environ.* 12 (2008) 345-351.
 - [8] G. Vernin, O. Merad, G.M.F. Vernin, R.M. Zamkotsian, C. Parkanyi, GC-MS analysis of *Artemisia herba-alba* Asso essential oils from Algeria, in: G. Charalambous (Ed.), *Food Flavors: Generation, Analysis and Process Influence*, Elsevier Science BV, Amsterdam, 1995, pp. 147-205. [http://dx.doi.org/10.1016/S0167-4501\(06\)80157-3](http://dx.doi.org/10.1016/S0167-4501(06)80157-3).
 - [9] A. Aafi, M.S. Taleb, M. Fechtal, Remarkable Species of the Flora of Morocco, CNRF, 2002, p. 146.
 - [10] A.M. Migahid, Flora of Saudi Arabia, 2nd ed., Riyadh University Press, Riyadh, Saudi Arabia, 1978, p. 61.
 - [11] S.M. Khazarji, L.A. Al-Shamony, H.A.A. Twaij, Hypoglycaemic effect of *Artemisia herba-alba*, *J Ethnopharmacol* 40 (1993) 163-166. [http://dx.doi.org/10.1016/0378-8741\(93\)90064-C](http://dx.doi.org/10.1016/0378-8741(93)90064-C).
 - [12] B. Imelouane, A. El Bachiri, M. Ankit, K. Khedid, J.P. Wathélet, H. Amhamdi, Essential oil composition and antimicrobial activity of *Artemisia herba-alba* Asso grown in Morocco, *Banats J Biotechnol* 1 (2010) 48-55.
 - [13] A.E-H.H. Mohamed, M.A. El-Sayed, M.E. Hegazy, S.E. Helaly, A.M. Esmail, N.S. Mohamed, Chemical constituents and biological activities of *Artemisia herba-alba*, *Rec Nat Prod* 4 (2010) 1-25.
 - [14] J.C. Chalchat, M.S. Gorunovic, Z.A. Maksimovic, S.D. Petrovic, Essential oil of wild growing *Mentha pulegium* L. from Yugoslavia, *Journal of Essential Oil Research* 12 (2000) 598-600.
 - [15] A. Zargari, Herbal Medicines, Publication of Tehran University, Iran, 1990, pp. 14-18.
 - [16] National Committee for Clinical Laboratory, Standards methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically, Wayne Pa., 1995, Vol. 15, p. 15 (approved standards No. M7-A3).
 - [17] H. Ismaili, L. Milella, S. Fkih-Tetouani, A. Ildrissi, A. Camporese, S. Sosa, et al., *In vivo* topical anti-inflammatory and *in vitro* antioxidant activities of two extracts of thymus satureioides leaves, *J. Ethnopharmacol* 91 (2004) 31-36.
 - [18] S. Bouhdid, Antimicrobial and antioxidant activities of essential oils: Biotechnological application to improve the quality of natural casings, *Journal of Applied Microbiology* 106 (5) (2009) 1558-1568.
 - [19] A. Akrou, Study of essential oils of some plants from the pastoral region of Matmata (Tunisia), *Institute of Arid Regions* 62 (1999) 289-292.
 - [20] M.M. Hudaib, T.A. Aburjai, Composition of the essential oil from *Artemisia herba-alba* grown in Jordan, *J Ess Oils Res* 18 (3) (2006) 301-304.
 - [21] S. Charchari, A. Dahoun, F. Bachi, A. Benslimani, *In vitro* antimicrobial of essential oils of *Artemisia herba-alba* and *Artemisia judacia* from Algeria, *Rivista-Italiana-EPPOS* 18 (1996) 3-6.
 - [22] I. Feuerstein, A. Danin, R. Segal, Constitution of the essential oil from an *Artemisia herba-alba* population of Spain, *Phytochem* 27 (2) (1988) 433-434.
 - [23] Z. Fleisher, A. Fleisher, R.B. Nachbar, Chemovariation of *Artemisia herba-alba* Asso. Aromatic plants of the Holy Land and the Sinai Part XVI, *J Ess Oils Res* 14 (2002) 156-160.
 - [24] G. Vernin, O. Merad, G.M.F. Vernin, R.M. Zamkotsian, C. Párkányi, GC-MS analysis of *Artemisia herba-alba* Asso essential oil from Algeria, *Dev Food Sci* 37A (1995) 147-205.
 - [25] D. Ouraini, A. Agoumi, M. Ismaili-Alaoui, K. Alaoui, Y. Cherrah, M.A. Alaoui, et al., Antifungal activity of oleic acid and of essential oils of *Thymus satureioides* L. et *Mentha pulegium* L., compared with antifungals in fungal dermatitis, *Phytotherapy* 1 (2007) 6-14.
 - [26] B. Chebli, M. Achouri, L.M. Idrissi Hassani, M. Hmamouchi, Chemical composition and antifungal activity of essential oils of seven Moroccan Labiatae against *Botrytis cinerea* Pers: Fr, *J. Ethnopharmacol* 89 (2003) 165-169.
 - [27] D. Ouraini, A. Agoumi, M. Ismaili-Alaoui, K. Alaoui, Y. Cherrah, M. Amrani, et al., Study of the activity of essential oils from aromatic plants with antifungal properties in different stages of development of dermatophytes, *Phytotherapy* 4 (2005) 147-157.
 - [28] M. Snoussi, H. Hajlaoui, E. Noumi, D. Usai, L.A. Sechi, S. Zanetti, et al., *In vitro* anti-vibrio spp. activity and chemical composition of some Tunisian plants, *World J. Microbiol. Biotechnol* 24 (2008) 3071-3076.
 - [29] H. Hajlaoui, N. Trabelsi, E. Noumi, M. Snoussi, H. Fallah, R. Ksouri, et al., Biological activities of the essential oils and methanol extract of two cultivated mint

- species (*Mentha longifolia* and *Mentha pulegium*) used in the Tunisian folkloric medicine, World J. Microbiol. Biotechnol 25 (2009) 2227-2238.
- [30] M. Mahboubi, G. Haghi, Antimicrobial activity and chemical composition of *Mentha pulegium* L. essential oil, J. Ethnopharmacol 119 (2008) 325-327.
- [31] E. Derwich, Z. Benziane, A. Boukir, GC/MS analysis and antibacterial activity of the essential oil of *Mentha pulegium* grown in Morocco, Research Journal of Agriculture and Biological Sciences 6 (3) (2010) 191-198.
- [32] N. Beghidji, N. Bouslimani, F. Benayache, S. Benayache, J.C. Chalchat, Composition of the oils from *Mentha pulegium* grown in different areas of the East of Algeria, Chem. Nat. Compd 43 (4) (2007) 481-483.
- [33] A. Remmal, T. Bouchikhi, K. Rhayour, M. Ettayebi, Improved method for the determination of antimicrobial activity of essential oils in agar medium, J. Essent. Oil Res 5 (1993) 179-184.
- [34] F. Şahin, M. Güllüce, D. Daferera, A. Sökmen, M. Sökmen, M. Polissiou, et al., Biological activities of the essential oils and menthol extract of *Origanum vulgare* ssp. Vulgare in the Eastern Anatolia region of Turkey, Food Control 15 (2004) 549-557.
- [35] J.C. Matasyoh, J.J. Kiplimo, N.M. Karubiu, T.P. Hailsstoks, chemical composition and antimicrobial activity of essential oil of *Tarchonanthus camphorates*, Food Chem 101 (2007) 1183-1187.
- [36] C.M. Mann, S.D. Cox, J.L. Markham, The outer membrane of *Pseudomonas aeruginosa* NCTC 6749 contributes to its tolerance to the essential oil of *Melaleuca alternifolia* (tea tree oil), Lett. Appl. Microbiol 30 (2000) 294-297.
- [37] C.J. Longbottom, C.F. Carson, K.A. Hammer, B.J. Mee, T.V. Riley, Tolerance of *Pseudomonas aeruginosa* to *Melaleuca alterifolia* (tea tree) oil is associated with the outer membrane and energy dependant cellular processes, J. Antimicrob. Chemother 54 (2004) 386-392.
- [38] G.Â.A. Pankey, L.Â.D. Sabath, Clinical relevance of *Bacteriostatic versus* bactericidal mechanisms of action in the treatment of gram positive bacterial infections, Clin Infect Dis 38 (2004) 64-70.
- [39] P.H. Andersen, N.J. Jensen, Mutagenic investigation of peppermint oil in the Salmonella/mammalian microsomal test, Mutat. Res 138 (1984) 17-20.
- [40] H. Oumzil, S. Ghoulami, M. Rhajaoui, A. Ildrissi, S. Fkih-Tetouani, M. Faid, et al., Antibacterial and antifungal activity of essential oils of *Mentha suaveolens* EHRH, Phytother. Res 16 (2002) 723-731.
- [41] D. Lopes-Lutz, D.S. Alviano, C.S. Alviano, P.P. Kolodziejczyk, Screening of chemical composition, antimicrobial and antioxidant activities of *Artemisia* essential oils, Phytochemistry 69 (8) (2008) 1732-1738.
- [42] S. Pattnaik, V.R. Subramanyam, M. Bapaji, C.R. Kole, Antibacterial and anti-fungal activity of aromatic constituents of essential oils, Microbios 89 (358) (1997) 39-46.