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**Abstract:** Salt is a major abiotic stress threatening crop plants such as rice. Tolerance to salt is complex and regulated by numerous genetic and non-genetic factors. To date most screens for salt tolerance rely on plant performance in stressed conditions. This article describes the ability to screen for salt tolerance in non-stressed conditions. The work is based on correlations between 62 rice genotypes in salt stress and non-stress conditions and measuring the intake of elements using Particle Induced X-ray Emission and X-ray Fluorescence. Roots and shoots were analysed though shoots provided easier and more robust materials to work with. Data were interpreted using multivariate statistical analysis which showed the intake ratio of elements across tolerant, moderately tolerant and susceptible rice genotypes. Tolerant genotypes exhibited a larger intake of elements, and a classification criteria based on Canonical Discriminant Analysis allowed differentiation of salt tolerant genotypes not only under salt stress but, significantly, also under non-stressed conditions. Thus, shoot element content in benign conditions can be used as an indicator for salt tolerance. These studies suggest that many salt tolerance mechanisms in rice cultivars are constitutive, they are not switched on by stress, and this has implications for physiological and genetic studies, especially in crop plants. The approach has practical application as it allows pre-screening in non-stressed conditions, from which candidate salt tolerant genotypes may be selected for subsequent testing in saline field conditions, selecting in benign conditions provides greater seed harvest of the next generation which can be used in multiple tests.

Key words: Rice, non-stress salt tolerance screening, canonical discrimination analysis, shoot element content, element ratios.

# **1. Introduction**

Soil salinity is prevalent in arid and semi-arid, and in coastal regions subject to inadequate irrigation and/or drainage. It is a major environmental constraint to crop productivity throughout the world [1-3], and especially problematic in irrigated agricultural systems. In general, salt tolerance in plants is associated with maintenance of growth and the ability to complete the life cycle. Salinity stress reduces growth and productivity [3, 4], and susceptible plants may die before reproduction. For crop plants salt tolerance may be generally defined as the ability to

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produce a yield in saline conditions.

Salt tolerance is often described as a complex trait, both genetically and physiologically [5]. In physiological terms salinity stress begins with low water potential in the root environment leading to a water deficit in the plant. This is followed by toxic effects of ions such Na<sup>+</sup> and Cl<sup>-</sup> on cells and tissues and results in a nutrient imbalance [6]. According to Munns and Tester [7] plants adapt to salinity through three distinct mechanisms: 1) osmotic adjustment, 2) ion exclusion and 3) tissue tolerance to the accumulation of toxic ions (particularly Na<sup>+</sup> or Cl<sup>-</sup>). Toxic elements often accumulate in older leaves, which is manifest by early senescence. When the death rate of older leaves is greater than the production of new leaves, the photosynthetic capacity drops and growth is retarded [7]. In salt tolerant mechanisms result the genotypes. these in maintenance of high shoot/root ratios and relatively high growth rates in saline conditions [6, 8, 9]. These complex physiological mechanisms often infer complex underlying genetics, which is problematic for plant breeders. Plant breeders are further constrained in that major genes, such developmental genes, e.g. those required by crops to grow in specific environments, often have pleiotropic effects on salt tolerance [10-12].

Rice is the most important cereal crop after maize in the world (http://faostat.fao.org/site/339/default.aspx) in terms of production. With respect to human consumption rice is pre-eminent; over half of the world's population depends on rice as a staple food, particularly in Asia and Africa [13, 14]. Fairhurst and Dobermann [15] reported that irrigated rice accounted for 55% of the global harvested area and contributed to 75% of global rice production. Rice is classified as a salt sensitive crop [16] and is particularly sensitive to salt stress at the seedling stage, less so at flowering and maturity [17]. Among rice genotypes there is a range of tolerance; three classes have been identified

(tolerant, moderate and susceptible) according to their response to salt stress [18]. The genetic variation for salt tolerance is of great interest as it provides potential for improvement through plant breeding.

Previous studies, based on traditional methods in assessing salt tolerance: root and shoot biomass production in salt treatments, have identified genotypes that can be used as standards for salt tolerance: tolerant, moderate and susceptible classes. Tolerance can be assessed using CID (Carbon isotope discrimination), which integrates plant performance/health over time and treatments. In our preliminary experiments, tissue element content was measured in six standard lines (two from each tolerance class) using atomic spectroscopy techniques (X-ray Fluorescence and Particle Induced X-ray Emission, a new application for these techniques). The data shows strong correlations with biomass production, which is a standard method for assessing salt tolerance. In order to validate these findings work was extended to a total of 62 rice genotypes from various regions and with known salt tolerances. Tests were carried out in saline and non-saline hydroponic conditions using a quick screening system developed by the International Rice Research Institute, IRRI [18] and modified by Plant Breeding and Genetics Laboratory PBGL [19].

# 2. Materials and Methods

# 2.1 Confirmation of Salt Tolerance in 6 Standard Genotypes Using Biomass Data

The six standard genotypes, 2 from each class (tolerant, moderate and susceptible) were checked for salt tolerance using root and shoot biomass data from hydroponics experiments in control and salt treatments. This confirmed the salt tolerance classes of the standard genotypes, which were then used in subsequent experiments, and provided information for optimum harvest data for biomass evaluation (16 days after salt treatment).

## 2.1.1 Plant Material

Six rice genotypes with known responses to salinity were used (data on the salt tolerance of these can be found in genotypes http://irri.org/our-science/genetic-diversity): two tolerant - 'Pokkali' and 'Nona Bokra', two of moderate tolerance - 'Bicol' and 'STDV', and two susceptible - 'IR29' and 'Taipei 309'. Genotypes 'Pokkali' and 'Nona Bokra' are both traditional tall, Indica rice cultivars from India. 'Bicol' is a relatively newly released moderately salt tolerant Indica cultivar derived from anther culture of an F<sub>1</sub> hybrid between IR5657-33-2 and IR4630-22-2-5-1-3 and 'STDV' is an anther culture derived semi-dwarf genotype from 'IR29' and classed as moderately tolerant. 'IR29' is a modern semi-dwarf Indica rice cultivar from IRRI, Philippines, and is susceptible to salinity; 'Taipei 309' is a Japonica cultivar also known to be susceptible to salinity. All six standard rice genotypes were obtained from IRRI.

## 2.1.2 Hydroponic Experiments

Hydroponic experiments were carried out in a glasshouse (temperature approx. day/night of: 30/20 °C  $(\pm 2 \text{ °C})$  with 50-70% relative humidity provided by misting). Seeds were pre-germinated in Petri dishes on filter paper with distilled water. Germinated seeds were transferred to mesh supports in contact with the surface of a hydroponic solution. After 2 weeks the seedlings were removed carefully and wrapped in sponge strips and transferred to test hydroponic tanks. A modified Yoshida nutrient solution was used in the hydroponic system [19, 20]. Two treatments were set up, a control (no added NaCl) and a salt treatment (NaCl, 10 dS/m or approximately 6.4 mg/L NaCl). These were applied after seedling establishment in hydroponics to four weeks old seedlings. Hydroponic solutions were replenished every 2-3 days. Details of the hydroponics protocol can be found at: http://www-naweb.iaea.org/nafa/pbg/public/manuals-p bg.html. Plant distribution was set up as a completely randomized design in the glasshouse with three

replications per treatment and ten plants per replication.

## 2.2 Element Composition Analysis

The six standard genotypes were analysed for element composition at 0, 12 and 16 days after salt treatment using atomic spectroscopy methods. Correlations were found with biomass and the element data could be used to discriminate between the three classes. A new finding was that patterns for element discrimination which were similar in control (no salt) and salt treatments, suggesting that salt tolerance could be evaluated in control conditions. 16 days proved to be the most informative harvest time for shoot element analysis.

## 2.2.1 Biomass Analysis

Plants of the six standard genotypes were sampled at 0, 4, 8, 12 and 16 days after treatment initiation (DAT). Growth parameters such as shoot and root biomass (fresh and dry weight), height and tiller number were used as a basis for determining salt tolerance. Roots of harvested plants were washed with water and gently blot-dried with paper tissue before recording fresh weight, plant height and number of tillers. Thirty plants were sampled *per* genotype *per* treatment, in total 360 plants *per* sampling time. For dry weight determination, shoots and roots were oven-dried at 70 °C for four days and then weighed.

2.2.2 Element Composition Analyses of Standard Genotypes

The biomass data were compared with element content data which were determined by X-ray Fluorescence spectrometer (XRF), see below. Shoots were sampled for each standard genotype at 0, 12 and 16 days after treatment initiation (DAT) using hydroponic tests as described above.

## 2.3 Validation Experiment

The study was extended to 62 rice genotypes of documented salt tolerance to validate the finding that element up-take in control conditions reflected

responses in salt stress conditions. Element data were compared with shoot biomass data and CID data.

## 2.3.1 Validation Experiment

62 rice genotypes with known responses to salinity were used as plant materials, this included the six standards. These comprised 23 tolerant, 13 moderately tolerant and 26 susceptible genotypes (Table 1). These genotypes were sourced mainly from IRRI, in addition to salt tolerance criteria; the rice genotypes were selected from a range of countries and include contemporary as well as traditional cultivars. Growth parameters such as shoot biomass (fresh and dry weights), height and tiller number were used as a basis for determining genotype performance under salinity. For dry weight determination, shoots and roots were oven-dried at 70 °C for four days and weighed. Then

the shoots samples were ground to fine powder for carbon isotope discrimination and elements contents by spectrometry.

2.3.2 Carbon Isotope Discrimination

CID has been used as a surrogate for salt tolerance in many studies [21-24]. Dried shoot samples of sixty genotypes from untreated and treated conditions were weighed and sealed into8x5 mm tin cups, then loaded into the auto-sampler of an EA (Elemental Analyzer) (Flash 2000, Thermo Scientific, Massachusetts, USA) coupled to an Isotope Ratio Mass Spectrometry (IRMS) (Isoprime, GV Instruments, Manchester, UK). The samples were flash combusted in a temporarily oxygen-enriched atmosphere of a combustion reactor (chromium oxide, silvered cobaltous/cobaltic oxide, quartz wool) held at 1,020 °C. The oxidation products

Table 1	Rice genotypes used,	their origin and lev	el of salt tolerance	(standards in bold).
				\ /

Susc	eptible	Moderate		Tolerant	
Genotype	Origin	BG 94-2	Sri Lanka	Genotype	Origin
BPT3402	India	DAMODAR	India	BHURA RATA 4-10	India
BR4	India	GETU	India	AKUNDO	Bangladesh
C5	United States	IR51491-AC5-1	IRRI	CHERIVIRUPPU	India
Giza 171	Egypt	POKKALI	Sri Lanka	IR 66946-3R-116-1-1	IRRI
HABA	Philippines	POKKALI (8558)	Sri Lanka	IR 66946-3R-149-1-1	IRRI
IET1444	India	IR 77660-B-9-1-3-2-1-7-5-1	IRRI	IR 58443-6B-10-3	Philippines
IR24	Philippines	IR 83460-4-B-4-2-1-1	IRRI	IRRI 147	IRRI
IR29	Philippines	IR 84115-10-B-AJY3-1-1	IRRI	IR 66946-3R-178-1-1	IRRI
M117	India	IR 84084-B-B-1-1	IRRI	IR11T189	IRRI
MADHUKAR	India	IR83420-B-AJY3-8-SDO1	IRRI	IR11T222	IRRI
SR 26B	Japan	Bicol (IAEA)	IRRI	NONA BOKRA	India
SUWON 143	Republic of Korea	STDV(IAEA)	IAEA	POKKALI 108921	India
KUATIK BENE	Indonesia	BG 94-2	Sri Lanka	TCCP 266-1-3B-13-1-3	IRRI
MK47-22	India	DAMODAR	India	HASAWI	Saudi Arabia
SINDANO	Kenya			IR 4630-22-2-5-1-3	IRRI
DHALIBORO 94	Bangladesh			CSR28	India
GASMAL 72-1	Bangladesh			AT 401	Sri Lanka
BINADHAN 7	Bangladesh			IR 72046-B-R-3	IRRI
BR 28	Bangladesh			CAPSULE	Bangladesh
BR 29	Bangladesh			A 69-1	Sri Lanka
NSIC Rc 222	Unknown			IR 55179-3B-11-3	IRRI
IR29	Philippines			NONA BOKRA (IAEA)	India
SADRI	Iran			POKKALI (IAEA)	Sri Lanka
TAIPEI 309 (IAEA)	Taiwan				
IR29 (IAEA)	Philippines				
NIPPON BARE	Japan				

were carried by a stream of helium through a reduction reactor (copper, quartz chips, quartz wool) at 650  $^{\circ}$ C. The resulting gases (primarily CO<sub>2</sub>, N<sub>2</sub> and H<sub>2</sub>O) were then carried through a magnesium perchlorate filter to remove water. The CO<sub>2</sub> and N<sub>2</sub> were separated in a packed chromatographic column, passed through a thermal conductivity detector, and carried into the source of the IRMS where the isotope ratios were measured against a pulse of reference gas of known isotopic composition. The carbon and nitrogen yields were estimated with a standard of known carbon and nitrogen content (laboratory standard S19) and calibration achieved from a regression of peak area versus the carbon and nitrogen content of the standard.

2.3.3 Element Composition

The elemental composition of the samples was analysed by XRF (X-ray Fluorescence) and PIXE (Particle Induced X-ray Emission). In total, fourteen elements were analysed: Na, Mg, P, S, Cl, K, Ca, Mn, Fe, Cu, Zn, Br, Rb and Sr. Shoot samples from three independent ten seedlings for each genotype were collected for both, control and exposed to salinity stress experiments. 0.5 g of milled shoots were pressed into a pellet (25 mm diameter) and prepared for both PIXE and XRF measurements.

Ten different certified RMs (Reference materials) of vegetable origin were used for calibration: brown bread (BCR191), cabbage (IAEA 359), hay (IAEA-V10), lichen (BCR 482), mixed polish herbs (INCT-MPH-2), olive leaves (BCR 62), orchard leaves (NBS1571), rye flour (IAEA-V8), spinach leaves (NBS1570A) and tea leaves (INCT-TL-1). Another RM of biological origin (milk powder, IAEA A11) was added to the list to enlarge the number of values for the concentration range of some elements, such as Cl, Mn, Cu and Rb.

For XRF, a spectrometer based on the use of a Pd-anode X-ray tube in combination with different secondary emission/scattering targets for excitation was used. Three excitation-measurement conditions were selected as to improve the X-ray production of different groups of elements as follows: Secondary Target (ST) of molybdenum (for Fe, Cu, Zn, Br, Rb, Sr); Cobalt ST (for K, Ca, Mn) and; HOPG (Highly Oriented Pyrolytic Graphite scatter element) for the excitation of low atomic number elements (Mg, P, S and Cl). Linear calibrations were made for quantification of each of the elements at the selected condition by measuring the RMs mentioned above. The calibrations were based on using selected energy radiation scattered at the sample as internal normalization of the measured fluorescent signal of element to compensate each for sample self-attenuation [25].

For PIXE, the measurements were made with a Silicon Drift Detector arranged in a configuration improved for the detection of low atomic number elements. The quantification was made based on determining individual geometry factors (H-values) for each element by measuring three different RMs. As the pellets cannot be considered as being electrically conductive, the effective charge deposited in the samples was determined by RBS (Rutherford Backscatter Spectrometry). The measured spectra were analysed using dedicated software (GUPIX for PIXE and SIMNRA for RBS). More details of XRF and PIXE can be found in Bado et al. [26].

2.3.4 Statistical Analysis

Data were recorded for P, K, Ca and Mg (macro-elements) and Fe, Mn, Zn, Cu, Cl, and Na (micro-elements) in shoot and root samples. In addition to the raw data, various salt tolerance indices were calculated including: K : Na, Ca : Na, Mg : Na, and Na : Cl ratios in shoots [27-33]. Mean values were evaluated at 5% significance level ( $P \le 0.05$ ) and in case of detection of significance different means were compared using Duncan's test.

The main steps followed for the multivariate statistical interpretation of the measured elemental concentrations were the following: (a) Multiplying the concentration values by the dry mass of the samples, in order to obtain the total amount of nutrients intake

by the plant; b) Transforming the resulting data to log 10 values, to avoid uneven concentration ranges of the elements that might impose different weights in analyses; (c) Reducing of subsequent the dimensionality of the data space using PCA (Principal Component Analysis) to ease the interpretation, and in the search for some ordination; (d) Establishing a classification criterion based on using two Canonical Discriminant Functions to differentiate three classes in the data set (tolerant, medium tolerance and susceptible varieties) and evaluation of group membership probabilities using CDA (Canonical Discriminant Analysis). The statistical interpretation was performed using the procedures included in the software SPSS 11.5.

# 3. Results

# 3.1 Confirmation of Salt Tolerance in Standard Genotypes

The biomass data were in accordance with the known salt tolerances of the 6 standard genotypes. The EC 10 dS/m NaCl salt stress treatment had a major effect on growth with respect to plant height, fresh and dry weight. However, no significant difference was recorded between genotypes for tiller number during the time course of the experiment. 'IR29' exhibited the biggest reduction in plant height at  $p \le 0.05$ . At 16 DAT both susceptible genotypes, 'Taipei 309' and 'IR29' were affected most by salinity

when compared with the salt tolerant genotypes, 'Pokkali' and 'Nona Bokra' (Table 2).

The least dry weight reduction percentage was observed in the tolerant genotype 'Nona Bokra' (16.4% and 7.1% respectively for shoot and root), and the greatest reduction was recorded for 'Taipei 309' (40.4% and 58.3% for shoot and root).

The salt treatment induced stationary or decreased biomass production with effects beginning to show differences among genotypes at 12 DAT (Fig. 1), as a consequence sampling at 0, 12 and 16 DAT were chosen for element compositional analysis by XRF. The tolerant genotypes 'Pokkali' and 'Nona Bokra' showed the least growth retardation (Fig. 1).

# 3.2 Element composition of standard genotypes

The element analysis of the 6 standard genotypes was used to study discrimination under salt stress and control treatments. Ratios of K, Ca, Mg and Cl with Na were associated with salt tolerance under treated than untreated condition (Table 3).

### 3.3 Validation Experiment

Element composition was determined using atomic spectroscopy and the data compared with biomass and CID measurements. Atomic spectroscopy was carried out using Particle Induced PIXE and XRF. Na, Mg, P, S and Cl were measured accurately with PIXE and Ca, Mn, Fe, Cu, Zn, Br, Rb and Sr were measured accurately

Table 2 Mean biomass data for each standard genotype at 16 DAT (NaCl, 10 dS/m).

Constants	Plant height	Whole plant fresh weight	Shoot dry weight	Root dry weight
Genotype 'Pokkali' 'Nona Bokra' 'Bicol' 'STDV' 'Taipei 309' 'IR29'	(cm)	(g)	(g)	(g)
'Daldrali'	80.33a	6.95 b	0.89 b	0.17 b
Роккап	[20.2]	[43.0]	plant fresh weightShoot dry weight (g)Root dry weight (g) $(g)$ $(g)$ $(g)$ $0.89$ b $0.17$ b $[31.5]$ $[20.3]$ $1.5$ a $0.26$ a $[16.4]$ $[07.1]$ $0.49$ c $0.10$ c $[28.0]$ $[31.8]$ $0.44$ cd $0.10$ c $[36.3]$ $[50.0]$ $0.16$ d $0.03$ d $[40.4]$ $[58.3]$ $0.31$ cd $0.06$ cd $[39.8]$ $[45.0]$	[20.3]
'None Polere'	86.99a	12.02a	1.5 a	0.26 a
INOIIA DOKIA	[21.3]	[29.0]	[16.4]	[07.1]
'Dical'	48.01b	3.33c	0.49 c	0.10 c
DICOI	[22.0]	[37.7]	[28.0]	[31.8]
(CTDV)	48.10b	3.08c	0.44 cd	0.10 c
SIDV	[27.0]	[55.0]	[36.3]	[50.0]
'Tainai 200'	42.47b	0.94 d	0.16 d	0.03 d
Taiper 509	[30.7]	[48.7]	[40.4]	[58.3]
(ID 20)	37.54 c	2.16cd	0.31 cd	0.06 cd
11/27	[31.9]	[47.6]	[39.8]	[45.0]

\*Means followed by same small letters denote no significant difference among genotypes at 10 dS/m salinity. [value]: % reduction compared to control.



Fig. 1 Shoot and root dry weight at 0, 4, 8, 12 and 16 DAT for the six standard rice genotypes from two treatments (0 and 10 dS/m). Each point represents the results of mean of 10 plants per genotype Tolerant genotypes: square symbol (solid: 'Pokkali' and open: 'Nona Bokra'); Moderate tolerant genotypes: Triangle symbol (solid: 'Bicol' and open: 'STDV'); Susceptible genotypes: round symbol (solid: 'Taipei 309' and open: 'IR29').

Table 3 J	Ratios of elements (H	K, Ca, Mg and	Cl) with Na in	the standard rice genoty	pes at 0 and 16 DAT under 10 dS/m
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Genotype 'Pokkali' 'Nona Bokra' 'Bicol' 'STDV'	Du	ration/Treatment		Mean element ratio			
Genotype	Day	EC (dS/m)	K:Na	Ca: Na	Mg:Na	Na:Cl	
	0	0	2.31 a	1.46 ab	1.87 c	0.51	
'Pokkali'	16	0	2.45 ab	1.44 ab	1.85 c	0.38 bc	
Genotype 'Pokkali' 'Nona Bokra' 'Bicol' 'STDV' 'Taipei 309' 'IR29'	16	10	1.32 a	1.42 a	0.51 a	0.47 bc	
	0	0	2.18 ab	1.70 a	3.02 a	0.52	
'Nona Bokra'	16	0	2.92 a	1.54 ab	2.31 b	0.27 c	
	10	10	1.20 ab	1.20 ab	Mean element ratioCa: NaMg:NaNa:Cl $1.46 ab$ $1.87 c$ $0.51$ $1.44 ab$ $1.85 c$ $0.38 bc$ $1.42 a$ $0.51 a$ $0.47 bc$ $1.70 a$ $3.02 a$ $0.52$ $1.54 ab$ $2.31 b$ $0.27 c$ $1.20 ab$ $0.69 a$ $0.28 c$ $1.25 ab$ $2.44 b$ $0.70$ $1.64 a$ $2.68 a$ $0.46 bc$ $0.79 abc$ $0.26 b$ $0.63 b$ $1.15 ab$ $2.74 ab$ $0.66$ $1.44 ab$ $2.41 ab$ $0.98 a$ $0.78 abc$ $0.20 b$ $0.65 b$ $1.06 b$ $2.41 b$ $0.98 a$ $0.38 c$ $0.07 b$ $1.01 a$ $1.38 ab$ $2.72 ab$ $0.95$ $1.43 ab$ $2.66 a$ $0.53 b$		
	0	0	1.70 abc	1.25 ab	2.44 b	0.70	
'Bicol'	16	0	2.15 bc	1.64 a	2.68 a	0.46 bc	
	16	10	0.57 bc	0.79 abc	NaNa:Cl6 ab $1.87 c$ $0.51$ 4 ab $1.85 c$ $0.38 bc$ 2 a $0.51 a$ $0.47 bc$ 0 a $3.02 a$ $0.52$ 4 ab $2.31 b$ $0.27 c$ 0 ab $0.69 a$ $0.28 c$ 5 ab $2.44 b$ $0.70$ 4 a $2.68 a$ $0.46 bc$ 9 abc $0.26 b$ $0.63 b$ 5 ab $2.74 ab$ $0.66$ 4 ab $2.41 ab$ $0.48 bc$ 8 abc $0.20 b$ $0.65 b$ 6 b $2.41 b$ $0.98 a$ 2 b $2.58 ab$ $0.98 a$ 8 ab $2.72 ab$ $0.95 a$ 3 ab $2.66 a$ $0.53 b$		
	0	0	1.53 bc	1.15 ab	2.74 ab	0.66	
'STDV'	16	0	1.92 bc	1.44 ab	2.41 ab	0.48 bc	
	16	10	0.60 bc	0.78 abc	0.20 b	0.65 b	
	0	0	1.31c	1.06 b	2.41 b	0.98	
'Taipei 309'	16	0	1.17 d	1.02 b	2.58 ab	0.98 a	
1	10	10	0.17 c	0.38 c	0.07 b	1.01 a	
	0	0	1.20 c	1.38 ab	2.72 ab	0.95	
'IR29'	17	0	1.77 c	1.43 ab	2.66 a	0.53 b	
	10	10	0.28 c	0.61 bc	0.09 b	b	

\*Ratios followed by the same letters for specific ratios (i.e. within columns) denote non-significant difference among genotypes in the same treatment.

with XRF analysis. The ratios of K, Ca, Mg and Cl with Na of the 62 rice genotypes were scattered among the salt tolerance. These ratios under treated and control (no NaCl) conditions did not show a clear differentiation of tolerant and susceptible genotypes. The data also showed the scattering of biomass performance of the 62 rice genotypes under the 10 dS/m NaCl. Therefore the elemental intake results were re-scaled to log-10 values, and the extracted PCA components were rotated using the Varimax method [34, 35], in order to provide greater agreement between axes and variable correlation, thus allowing a better interpretation of the observed differences in elemental contents due salt treatments. The coefficients of each of the original variables (elemental concentration) in extracted principal components (principal component loadings) provide information for the identification of correlated variables and their contribution to the variability in the data set.

The tendency in the variations of concentrations of the fourteen elements in the different genotypes was explored for the two data sets containing the control and exposed to salinity stress samples, respectively. The results of the PCA are summarized in Tables 4 and 5, and revealed that the two first components accounted for 65 % and 23 % of variability of the data set, respectively (Table 4). The largest variability in the data set, nearly 65 % is accounted for by the nutrient elements (see larger loading values in Table 4), even larger than the contribution to variability due to the elements related to the salinity stress (Na, Cl and Br). The tendency in variations of the elemental mass fractions with salt status and treatment can be observed from Fig. 2.

Each sample is represented in Fig. 2 as occupying a location by its principal component scores in the 2D space formed by the first extracted principal components. As samples are labelled according to their tolerance and the exposure to salt treatment at which the plant tissue was collected, it can be observed from this ordination that there is a marked difference in the composition of the samples. The ordination of the samples in the PC space reveals two groups corresponding to the exposure to salt levels: treated samples appear clustered in the upper left area of the graph whereas untreated appear in the lower right area. The results obtained for the samples that underwent saline treatment show that there is a decrease in the intake of nutrients Mg, P, S, K, Ca, Mn, Fe, Cu and Zn compared to the control (non-salt treated) samples. The decrease is more pronounced for the susceptible genotypes, whereas tolerant ones exhibit a larger intake of these elements. All elements showed a decrease under salt stress except for Na and Cl which accumulated in the shoots.

PCA analysis (Fig. 2) showed differentiation between the treatments and genotype tolerance to salinity.

		Initial eiger	nvalues	Extraction sums of squared loadings			Rotation sums of squared loadings		
Component	Total	% of variance	Cumulative %	Total	% of variance	Cumulative %	Total	% of variance	Cumulative %
1	7.854	65.452	65.452						
2	2.734	22.784	88.236						
3	.489	4.074	92.310						
4	.204	1.697	94.007						
5	.159	1.322	95.329						
6	.135	1.126	96.455	7.854	65.452	7.808	7.808	65.068	65.068
7	.108	.903	97.359	2.734	88.236	2.780	2.780	23.168	88.236
8	.098	.819	98.178						
9	0.78	.650	98.828						
10	.064	.536	99.364						
11	.052	.434	99.798						
12	.024	.202	100.000						

 Table 4
 Eigenvectors and percent of variance explained the variation in principal component analysis.

		Raw		Rescaled			
Element		Component		Component			
	1	2	1	2			
L_TK	.243	024	.964	093			
L_TS	.193	009	.960	046			
L_TFE	.225	003	.946	014			
L_TP	.171	.019	.933	.106			
L_TMG	.208	007	.932	031			
L_TCU	.158	.027	.921	.156			
L_TZN	.158	.032	.913	.182			
L_TCA	.137	.037	.890	.243			
L_TMN	.156	001	.784	004			
L_TCL	.047	.268	.166	.955			
L_TNA	246	.680	340	.939			
L_TBR	.062	.215	.257	.893			







Fig. 2 Ordination of the samples according to their scores in the PC-space. Each point represents the results for individual genotype samples, which is in turn labelled according to the tolerance to salt (S - susceptible, M - Moderate, T- tolerant genotypes) and to the group C - Control, S - Salt treated. Note that the pattern for control genotype (closed symbols) shows a strong similarity to the pattern of salt treatment data (open symbols).

These clusterings were less pronounced for root data in the analysis of standard genotypes. As a consequence the untreated shoot data set from validation experiment was explored for differentiation under control condition in a CDA.

These observations support the feasibility of classifying the rice genotypes into three tolerance classes, based on the contents of elements in seedling shoots. Linear discriminant analysis is a method commonly used in statistics to find linear combination of property values allowing separating two or more classes of objects. Canonical discriminant analysis aims to find linear and uncorrelated functions defining a k-l canonical functions that allow achieving a better separation of k expected classes. CDA also allows calculating the probability of belonging to a particular class.

The results of such interpretation of the results obtained for the entire population of control shoot samples are represented in Fig. 3. The probability of belonging to its assumed class was found to be more than 75% for the majority of tolerant and susceptible genotypes, over 80% of all cases. The canonical analysis allowed the three salt tolerant classes

(tolerant, moderate tolerant and susceptible) to be separated.

The element composition data were compared to CID. CID in shoot tissues at 16 DAT (16 DAT was chosen as it gave the best salt tolerance differentiation for biomass). The CID plotted with the dry matter revealed significant differences between untreated and treated shoot samples and genetic variability among the rice genotypes (Fig. 4). The value of CID ranged from -30.86 to -28.73 and -30.19 to -27.64 respectively for untreated and treated shoot indicating that under salt stress CID became more positive. Obtained results allowed the clustering of untreated and treated samples with quite clear differentiation of the tolerant, moderate tolerant and susceptible genotypes.



Fig. 3 Clustering of rice genotypes with known salt tolerance into the three salt tolerant classes based on conical discrimination analysis of 16 DAT shoot element content. The mid-point for each cluster is indicated by a solid symbol. Each open symbol represents the mean of three replications of ten plants *per* individual genotype samples, which is in turn labelled according to the tolerance to salt. Triangle symbol: susceptible, square symbol: moderate, round symbol: tolerant genotypes).



Dry weight per plant (g)

Fig. 4 Relationship between CID and shoot dry matter showing the clustering of untreated and treated samples. Each point represents the results for individual genotype samples, which is in turn labelled according to the tolerance to salt (susceptible: round symbol, moderate: triangle symbol and tolerant genotypes: square symbol) and to the group Control (open symbols), treated (solid symbols).

# 4. Discussion

Rice is a crop plant that is relatively susceptibility to salinity [17, 36], but genotypic differences are known at different growth stages [37]. The current work used a hydroponic system to study seedling response (the most susceptible growth stage) of 62 rice genotypes of varying tolerance to salt. Various growth parameters were measured in the initial experiments. The number of tillers was found to be of no value as an indicator of salt tolerance in rice at the seedling stage as all genotypes behaved the same in this respect, despite the fact that Nicolas et al. [38] reported in wheat that salinity may inhibit tiller formation and even tiller abortion during tiller emergence. Differences between standard genotypes were observed for plant height and shoot weight (fresh and dry) between treatments. 10 dS/m NaCl salt affected rice seedling growth showing significant decreased of the shoot fresh and dry weight, plant height (Table 1 and Fig. 1). 'Pokkali' and 'Nona Bokra' are both tall genotypes whereas moderate and susceptible are semi-dwarfs. Tall types were therefore associated with salt tolerance; the height factor could be removed by calculating height reduction in salt stress as compared to control conditions for all genotypes. 'Pokkali' and 'Nona Bokra' were used in experiments by Banik et al. [39] who observed growth reduction after 21 days of 10 dS/m salt stress of 20% and 21% after 16 days, respectively. The salt tolerance of 'Pokkali' and 'Nona Bokra' has been reported

independently by several workers [19, 40-43]. Our data corroborate these findings. More susceptible genotypes resulted in great reductions in plant height (Table 1, Fig. 1).

The 62 rice genotypes studied with known salt tolerance exhibited diversity with respect to plant vigour. Thus a range of plant heights were recorded: tall, semi-dwarf and dwarf. Each salt tolerant category classes within tall, semi-dwarf and dwarf genotypes while, tall and tolerant genotypes record similar performance as well as semi-dwarf or dwarf tolerant genotypes. The performance under salt is independent of the plant tallness. Thus, different performance was observed regarding the fresh and dry weight and plant height even after normalization which was not always in accordance with the reported salt tolerance.

Carbon isotope discrimination (CID) was generally effective in classifying salt tolerant and salt susceptible genotypes with tolerant genotypes having a tendency to maintain more negative  $\delta^{13}$ C values compared to the susceptible ones. CID has been used as an indirect screening method for selecting better and adapted cultivars to adverse environmental conditions such as drought, salt, etc. Many studies have used CID in selecting salt tolerance in rice [21-24, 44, 45]. Our CID data are in agreement with the salt tolerance of the standard lines and confirmed their classes as tolerant, moderately tolerant and susceptible. For more information on the use CID as a surrogate for salt tolerance, the reader is referred to recent publications [46-48]. Here we focus on the ability to screen for salt tolerance using element composition in non-stressed conditions.

Element analyses were carried out on 12 elements, but one objective was to streamline analyses to include only the most informative elements in developing a simplified salt tolerance analysis. Leaf samples were found to be more informative and easier to sample than roots. Combined analyses of shoots were performed on the elements with the greatest effects (Na, Cl, Mg, K and P) in salt tolerance tests on the six standard genotypes (Table 3). This was not confined to total content of these elements in shoots, but also included informative ratios, such as K:Na, CaNa, Mg:Na and Na:Cl. These element ratios yielded the same discrimination between the rice genotypes as total elements in PCA and CDA analysis. K:Na, Ca:Na, and Mg:Na were positively correlated with salt tolerance (Table 3). Increases in Na and Cl contents of shoots were associated with salt susceptibility, but the effect of Cl was weak, suggesting that Na is the main toxic element for all genotypes tested. However, damaging effects of Cl are observed with moderate and susceptible genotypes. The toxic effects of Na are well known and correlated with the grain yield under saline conditions [49-53], whereas there are few reports on toxic effects of Cl in salinity experiments [3, 9, 33, 54-58]. For further information on the roles of Na and Cl in plants under salt stress see Teakle and Tyerman [59] and Flowers et al. [60].

Single or combined element ratios were scattered in the 62 rice genotypes under control and salt stress conditions within each salt category compared to the standards. The data are in agreement with the findings of Chunthaburee et al. [61] who used 12 rice cultivars including two of the standards used here, 'Pokkali' and 'IR29', in which K/Na ratio was only able to discriminate salt tolerance under salt stress conditions. This demonstrates the complexity of mechanism involved in salt tolerance and the difficulty in identifying a single criteria, see also Ashraf [62] and, Shahbaz and Ashraf [63]. Our data show that salt treatment affects the composition of several elements. However, the loading of each element may differ from others because of its importance in various physiological systems, plant growth, regulations, etc. (Tables 4 and 5). In order to get a handle on these multiple effects, the fourteen elements detected and analysis by PIXE and XRF were combined by PCA and CDA. The resulting clustering of the genotypes into 3 classes provides evidence of genetic variation

for salt tolerance in rice in respect to shoot accumulation and discrimination of elements (Figs. 2 and 3). Furthermore the same trend of clustering and differentiation was observed with element content under control and saline conditions (Figs. 2 and 3). This is a new and interesting finding. The contribution of different elements in the PCA show that shoot Na, Cl, Mg, K and P concentrations exhibit big changes between treatments (Tables 4 and 5). The results obtained for the control samples revealed that tolerant genotypes have comparatively larger intakes of elements than moderate-tolerant and susceptible ones. Thus the ability to discriminate elements is important feature of salt tolerant genotypes [64]. The clustering in response to salt stress is independent of plant type (tall, semi-dwarf and dwarf) as these were found in each salt category. Thus the linkage between salt tolerance and height can be broken, an important point for plant breeders.

Screening for salt tolerance in control conditions presents many advantages for plant breeders. An initial non-destructive screen for salt tolerance may be carried out in benign, fertile conditions on young plants. Early generations may be tested and grown on to produce relatively large numbers of seed for subsequent field testing in saline conditions for confirmation. In addition that allows further screening for other interested traits. These findings may be applicable to other crops.

## **5.** Conclusions

Salt tolerance in rice cultivars can be screened for in non-stressed conditions. This conclusion was formulated from a series of experiments in which the salt tolerance of six standards was confirmed using biomass data from hydroponic culture in saline and non-saline conditions. Element composition was determined by PIXE and XRF, which is a new application for these atomic spectroscopy techniques. Element composition data were able to discriminate between the six standard rice genotypes classed as tolerant, moderate or susceptible to salt (two in each class). Furthermore, the ranking of salt tolerance with respect to element composition was the same in salt and control treatments. These findings were validated by extending the study to a large range (62) of rice genotypes from various countries.

The concentration of elements in rice shoots can serve as a basis for predicting the response to salinity stress. The most effective measurements are shoot dried weight and elemental composition, particularly the ratio Ka:Na and Mg:Na under salt stress taken at 16 DAT. That may be assessed from leaf biopsies and plants possessing element compositions indicative of salt tolerance selected. Testing for salt tolerance in benign conditions offers a simple, non-destructive pre-screen for plant breeders and since the tests may be conducted in fertile areas large amounts of seed may be harvested for subsequent testing, e.g. in saline fields.

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The authors declare no conflict of interest.

## References

 Rengasamy, P. 2010. "Soil Processes Affecting Crop Production in Salt-Affected Soils." *Funct. Plant Biol.* 37: 613-20.

- [2] Islam, M. S., Rahman, M. A., Sultana, N., Nath, B. and Paul, A. 2012. "Using Geospatial Techniques to Assess the Salinity Impact on Agricultural Landuse: a Study on Shyamnagar Upazila, Satkhira." *JAEID* 106 (2): 157-69.
- [3] Munns, R., James, R. A., Xu, B., Athman, A., Conn, S. J. and Jordans, C. et al. 2012. "Wheat Grain Yield on Saline Soils is Improved by an Ancestral Na<sup>+</sup> Transporter Gene." *Nat Biotechnol* 30: 360-4.
- [4] Horie, T., Karahara, I. and Katsuhara, M. 2012. "Salinity Tolerance Mechanisms in Glycophytes: An Overview with the Central Focus on Rice Plants." *Rice* 5: 11-28.
- [5] Flowers, T. J. and Flowers, S. A. 2005. "Why Does Salinity Pose Such a Difficult Problem for Plant Breeders?" *Agr Water Manage* 78: 15-24.
- [6] Mansour, M. M. F., Salama, K. H. A., Ali, F. Z. M. and Hadid, A. F. A. 2005. "Cell and Plant Responses to NaCl in *Zea mays* L. Cultivars Differing in Salt Tolerance." *Gen Appl. Plant Physiol.* 31: 29-41.
- [7] Munns, R. and Tester, M. 2008. "Mechanisms of Salinity Tolerance." *Annu Rev Plant Biol.* 59: 651-81.
- [8] Munns, R., Husain, S., Rivelli, A. R., James, R. A, Condon, A. G. and Lindsay, M. P. et al. 2002. "Avenues for Increasing Salt Tolerance of Crops, and the Role of Physiologically Based Selection Traits." *Plant and Soil* 247: 93-105.
- [9] Munns, R. 2002. "Comparative Physiology of Salt and Water Stress." *Plant Cell and Environment* 25: 239-50.
- [10] Andeden, E. E., Yediay, F. E., Baloch, F. S., Shaaf, S., Kilian, B., Nachit, M. M. and Zkan, H. 2011.
  "Distribution of Vernalization and Photoperiod Genes (Vrn-A1, Vrn-B1, Vrn-D1, Vrn-B3, Ppd-D1) in Turkish Bread Wheat Cultivars and Landraces." *Cereal Res Commun.* 39: 352-64.
- [11] Eagles, H. A., Cane, K., Trevaskis, B., Vallance, N., Eastwood, R. F. and Gororo, N. N. et al. 2014. "Ppd1, Vrn1, ALMT1 and Rht Genes and Their Effects on Grain Yield in Lower Rainfall Environments in Southern Australia." *Crop Pasture Sci.* 65: 159-70.
- [12] Kiss, T., Balla, K., Veisz, O., Láng, L., Bedő, Z. and Griffiths, S. et al. 2014. "Allele Frequencies in the VRN-A1, VRN-B1 and VRN-D1 Vernalization Response and PPD-B1 and PPD-D1 Photoperiod Sensitivity Genes, and Their Effects on Heading in a Diverse Set of Wheat Cultivars (*Triticum aestivum* L.)." *Mol Breed* 34 (2): 297-310.
- [13] Sankar, P. D., Maam, S. and Selvaraj, C. I. 2011. "Rice Breeding for Salt Tolerance." *Res Biotechnol* 2 (2): 1-10.
- [14] Todaka, D., Nakashima, K., Shinozaki, K. and Yamaguchi-Shinozaki, K. 2012. "Toward Understanding Transcriptional Regulatory Networks in Abiotic Stress Responses and Tolerance in Rice." *Rice* 5: 6-14.
- [15] Fairhurst, T. H. and Doberman, A. 2002. "Rice in the

Global Food Supply." Better Crop International 16: 3.

- [16] Shannon, M. C., Rhoades, J. D., Draper, J. H., Scardaci, S. C. and Spyres, M. D. 1998. "Assessment of Salt Tolerance in Rice Cultivars in Response to Salinity Problem in California." *Crop Sci.* 38 (2): 394-8.
- [17] Roy, S. J., Tucker, E. J. and Tester, M. 2011. "Genetic Analysis of Abiotic Stress Tolerance in Crops." *Curr Opin Plant Biol* 14: 232-9.
- [18] Gregorio, G. B., Senadhira, D. and Mendoza, R. T. 1997."Screening Rice for Salinity Tolerance." IRRI Discussion Paper Ser 22. Manila.
- [19] Afza, R., Zapata-Arias, F. J., Zwiletitsch, F., Berthold, G. and Gregorio, G. 1999. "Modification of a Rapid Screening Method of Rice Mutants for NaCl Tolerance Using Liquid Culture." *Mutat Breed Newsl* 44: 25-28.
- [20] Yoshida, S., Forno, D. A., Cock, J. H. and Gomez, K. A. 1976. "Laboratory Manual for Physiological Studies of Rice." Las Banos, Laguna: IRRI, 83.
- [21] Shaheen, R. and Hood-Nowotny, R. C. 2005. "Carbon Isotope Discrimination: Potential for Screening Salinity Tolerance in Rice at the Seedling Stage Using Hydroponics." *Plant Breed* 124: 220-4.
- [22] Dadkhah, A. and Ghorbanzadeh-Neghab, M. 2012. "Carbon Isotope Discrimination, A Tool for Screening of Salinity Tolerance of Genotypes. *International Scholarly* and Scientific Research & Innovation 6 (9): 121-4.
- [23] Brugnoli, E. and Lauteri, M. 1991. "Effects of Salinity on Stomatal Conductance, Photosynthetic Capacity, and Carbon Isotope Discrimination of Salt-Tolerant (*Gossypium hirsutum* L.) and Salt-Sensitive (*Phaseolus vulgaris* L.) C (3) Non-Halophytes." *Plant Physiol* 95 (2): 628-35.
- [24] Shaheen, R. and Hood-Nowotny, R. C. 2004. "Effect of Drought and Salinity on Carbon Isotope Discrimination in Wheat Cultivars." *Plant Sci* 168: 901-9.
- [25] Padilla, R., van Espen, P. and Torres, P. P. G. 2006. "The Suitability of XRF Analysis for Compositional Classification of Archaeological Ceramic Fabric: a Comparison with a Previous NAA Study." *Anal Chim Acta* 558: 283-9.
- [26] Bado S., Padilla-Alvarez R., Migliori A., Forster B.P., Jaksic M., Diawara Y., Kaiser R.B. and Laimer M. 2016. "The Application of XRF and PIXE in the Analysis of Rice Shoot and Compositional Screening of Genotypes." Nuclear Instruments and Methods in Physics Research Section B: Beam Interactions with Materials and Atoms, 371: 407-12.

http://dx.doi.org/10.1016/j.nimb.2015.08.081.

- [27] Tal, M. 1985. "Genetics of Salt Tolerance in Higher Plants: Theoretical and Practical Considerations." *Plant* and Soil 89: 199-226.
- [28] Dasgan, H. Y., Aktas, H., Abak, K. and Cakmak, I. 2002.

"Determination of Screening Technique to Salinity Tolerance in Tomatoes and Investigation of Genotypes Responses." *Plant Sci.* 163: 695-703.

- [29] Hussain, N., Ali, A., Khan, A. G., Rehman, O-U. and Tahir, M. 2003. "Selectivity of Ions Absorption as Mechanism of Salt Tolerance in Rice [variety Shaheen Basmati]." *Asian J Plant Sci.* 2: 445-8.
- [30] Flowers, T. J. 2004. "Improving Crop Salt Tolerance." J Exp Bot 55: 307-19.
- [31] Moradi, F. and Ismail, A. M. 2007 "Responses of Photosynthesis, Chlorophyll Fluorescence and ROS-Scavenging Systems to Salt Stress during Seedling and Reproductive Stages in Rice." *Ann Bot* 99: 1161-73.
- [32] Saleh, B. 2011. "Effect of Salt Stress (NaCl) on Biomass and K<sup>+</sup>/Na<sup>+</sup> Ratio in Cotton." *J stress Physiol Biochem.* 7 (4): 5-14.
- [33] Saleh, B. 2011. "Ion Partitioning and Mg<sup>2+</sup>/Na<sup>+</sup> Ratio under Salt Stress Application in Cotton." *J Stress Physiol Biochem* 7 (4): 292-300.
- [34] Mardia, K. V., Kent, J. T. and Bibbly, J. M. 1979. "Multivariate Analysis." London: Academic Press Inc.
- [35] Johnson, R. A. and Wichern, D. W. 2007. "Applied multivariate statistical analysis." Sixth Eds. Prentice-Hall, Englewood Cliffs, USA.
- [36] Kavitha, P. G., Miller, A. J., Mathew, M. K. and Maathuis, F. J. M. 2012. "Rice Cultivars with Differing Salt Tolerance Contain Similar Cation Channels in Their Root Cells." *J Exp Bot* 63 (8): 3289-96.
- [37] Zeng, L., Shannon, M. C. and Grieve, C. M. 2002.
   "Evaluation of Salt Tolerance in Rice Genotypes by Multiple Agronomic Parameters." *Euphytica* 127: 235-45.
- [38] Nicolas, M. E., Munns, R., Samarakoon, A. B., Gifford, R. M. 1994. "Elevated CO<sub>2</sub> Improves the Growth of Wheat under Salinity." *Aust J Plant Physiol* 20: 349-60.
- [39] Banik, M., Karim, N. H. and Haque, M. Z. 1994. "Salinity Tolerance of Rice as Related to Growth and Physiological Characteristics. *Ann Bangladesh Agric* 4: 41-6.
- [40] Flowers, T. J. and Yeo, A. R. 1981. "Variability in the Resistance of Sodium Chloride Salinity within Rice [Oryza sativa L.] Varieties." New Phytol 88: 363-73.
- [41] Akita, S. and Cabuslay, G. S. 1990. "Physiological Basis of Differential Response to Salinity in Rice." *Plant and Soil* 123: 277-94.
- [42] Yeo, A. R., Yeo, M. E. and Flowers, T. J. 1990. "Screening of Rice [*Oryza sativa* L.] Genotypes for Physiological Characters Contributing to Salinity Resistance, and Their Relationship to Overall Performance." *Theor Appl Genet* 79: 377-84.
- [43] Dionisio-Sese, M. L. and Tobita, S. 1998. "Antioxidant Responses of Rice Seedlings to Salinity Stress." *Plant Sci.*

135: 1-9.

- [44] Islam, M. M., Begum, S. N., Emon, R. M., Halder, J. and Manidas, A. C. 2011. "Carbon Isotope Discrimination in Rice under Salt Affected Conditions in Bangladesh". In Greater Agronomic Water Use Efficiency in Wheat and Rice Using Carbon Isotope Discrimination. IAEA-TECDOC-1671, 7-23.
- [45] Ismail, A. M., Katimbang, M. L., Egdane, J. A. and Thomson, M. J. 2011. "Carbon Isotope Discrimination and Salt Tolerance in Rice." In *Greater Agronomic Water* Use Efficiency in Wheat and Rice Using Carbon Isotope Discrimination. IAEA-TECDOC-1671, 49-66.
- [46] Cui, Y. Q., Ma, J. Y. and Sun, W. 2011. "Application of Stable Isotope Techniques to the Study of Soil Salinization." *J Arid Land* 3 (4): 285-91.
- [47] Dadkhah, A. 2013. "Effect of Salinity on Carbon Isotope Discrimination of Shoot and Root of Four Sugar Beet (Beta vulgaris L.) Cultivars." J Agric Sci. Technol (15): 901-10.
- [48] Ivlev, A. A., Pichouzkin, V. I., Tarakanov, I. G. 2013. "Soil Salinity Effect on Carbon Isotope Composition of Plant Biomass." *Adv St Biol* 5 (5): 223-34.
- [49] Maathuis, F. J. M. and Amtmann, A. 1999. "K<sup>+</sup> Nutrition and Na<sup>+</sup> Toxicity: The Basis of Cellular K<sup>+</sup>/Na<sup>+</sup> Ratios." *Ann. Bot* 84: 123-33.
- [50] Zeng, L., Poss, J. A., Wilson, C., Draz, A. E., Gregorio, G. B. and Grieve, C. M. 2003. "Evaluation of Salt Tolerance in Rice Genotypes by Physiological Characters." *Euphytica* 129: 281-92.
- [51] Shereen, A., Ansari, R. U., Yasmin, S., Raza, S., Mumtaz, S., Khan, M. A. and Mujtaba, S. M. 2007. "Physiological Responses of Rice [Oryza sativa L.] to Saline Stress." *Pak J Bot* 39 (7): 2527-34.
- [52] Khan, M. A., Shirazi, M. U., Khan, M. A., Mujtaba, S. M., Islam, E. and Mumtaz, S. et al. 2009. "Role of Proline, K/Na Ratio and Chlorophyll Content in Salt Tolerance of Wheat [Triticum aestivum L.]." *Pak J Bot* 41: 633-8.
- [53] Zhang, J.-L., Flowers, T. J. and Wang, S.-M. 2009. "Mechanisms of Sodium Uptake by Roots of Higher Plants." *Plant Soil* 326: 45-60.
- [54] Ashraf, M. and McNeilly, T. 1987. "Salinity Effects on Five Cultivars/Lines of Pearl Millet (Pennisetum americanum [L] Leeke)." *Plant and Soil* 103: 13-9.
- [55] Sharma, S. K. 1996. "Effects of Salinity on Uptake and Distribution of Na<sup>+</sup>, Cl<sup>-</sup> and K<sup>+</sup> in Two Wheat Cultivars." *Biol Plantarum* 38 (2): 261-7.
- [56] Chartzoulakis, K., Loupassaki, M., Bertaki, M. and Androulakis, I. 2002. "Effect of NaCl Salinity on Growth, Ion Content and CO<sub>2</sub> Assimilation Rate of Six Olive Cultivars." *Scientia Horticulturae* 96: 235-47.
- [57] Akhtar, J., Saqib, Z. A., Saleem, I. and Haq, M. A. 2010.

"Evaluating Salt Tolerance Cotton Genotypes at Different Levels of NaCl Stress in Solution and Soil Culture." *Pak J Bot* 42: 2857-66.

- [58] Keutgen, A. J. and Pawelzik, E. 2009. "Impacts of NaCl Stress on Plant Growth and Mineral Nutrient Assimilation in Two Cultivars of Strawberry." *Environ Exp Bot* 65: 170-6.
- [59] Teakle, N. L. and Tyerman, S. D. 2010. "Mechanisms of Cl<sup>-</sup> Transport Contributing to Salt Tolerance." *Plant Cell* and Environ 33: 566-89.
- [60] Flowers, T. J., Munns, R. and Colmer, T. D. 2014. "Sodium Chloride Toxicity and the Cellular Basis of Salt Tolerance in Halophytes." *Ann Bot* 115 (3): 327-31.
- [61] Chunthaburee, S., Dongsansuk, A., Sanitchon, J., Pattanagul, W. and Theerakulpisut, P. 2015.
  "Physiological and Biochemical Parameters for Evaluation and Clustering of Rice Cultivars Differing in Salt at Seedling Stage. *Saudi J Biol Sci.*
- [62] Ashraf, M. 2004. "Some Important Physiological Selection Criteria for Salt Tolerance in Plants." *Flora* 199: 361-76.
- [63] Shahbaz, M. and Ashraf, M. 2013. "Improving Salinity Tolerance in Cereals." *Crit Rev Plant Sci.* 32 (4): 237-49.
- [64] Zeng, L. 2005. "Exploration of Relationship between Physiological Parameters and Growth Performance of Rice [*Oryza sativa* L.]." *Plant and Soil* 268: 51-9.